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POLYANION INHIBITORS OF HUMAN IMMUNODEFICIENCY VIRUS. Part IV. - POLYMERIZED ANIONIC SURFACTANTS: INFLUENCE OF THE DENSITY AND DISTRIBUTION OF ANIONIC GROUPS ON THE ANTIVIRAL ACTIVITY.

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Abstract: The synthesis of a series of new polyanions, via γ -polymerization, of ω -unsaturated multi-charged anionic surfactants, has been realized. These polyanions were evaluated for their activity against HIV-1. All tested compounds proved active without presenting toxicity to the host cells (CEM-4 or MT-4).

Various polyanions have proven inhibitory to HIV replication; their activity has been attributed to interference with virus adsorption, due to a shielding of the viral envelope glycoprotein gp120^{1,2}.

Recently, we have shown that polyanions obtained by γ-polymerization in micellar solutions of ω-unsaturated anionic surfactants (Figure 1a) with the polar head derived either from simple anionic groups³, aminoacids or dipeptides⁴, or saccharidic residues⁵, markedly inhibit the replication of HIV-1 and HIV-2 in cell culture. All starting monomeric surfactants were monoanionic compounds.

In a recent article², it has been postulated that the antiviral potency and activity spectrum of polyanions could depend upon the density and the distribution of the negative charges. This hypothesis prompted us to explore the antiviral activities of polyanions obtained by γ -polymerization of ω -unsaturated multi-charged anionic surfactants, i.e. bi-, tri- or tetra-charged monomers (Figure 1b, c and d).

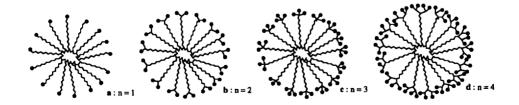


Figure 1.- Schematic structure of polyanions obtained by γ -polymerization of ω -unsaturated n-charged anionic surfactants (\bullet : mono-anionic group).

In this work, we present the synthesis of a series of polyanions that originate from monomers of a C₁₁ linear chain bearing a carbon-carbon double bond at one end, and a multi-charged anionic head at the other end. Antiviral activities of these compounds are also presented.

398 A. LEYDET et al.

Syntheses

Except for the compound 3m resulting from the malonic acid alkylation, all surfactants are amide derivatives obtained from natural or modified α -amino-acids, or from tris-(hydroxymethyl) aminomethane.

Dianionic compounds

1 - Compounds derived from malonic acid: the sodium 2-(undec-10-enyl)propanedioate 2 was obtained according to a classic malonic synthesis⁶. The condensation of the 11-bromoundec-10-ene 1³ on the diethylmalonate was realized in the presence of sodium ethoxide to give diester 2, which was saponified by sodium hydroxyde in water/acetone solution to give the surfactant 3m (Scheme 1) in 90% yield⁷.

Scheme 1
$$(CH_2)_9$$
-Br CO_2 Et CO_2 Et CO_2 Et CO_2 Et CO_2 Et CO_2 Et CO_2 INa⁺ CO_2 INa⁺ CO_2 INa⁺ CO_2 INa⁺

2 - Compounds derived from L-aspartic and L-glutamic acids: ω-unsaturated surfactants derived either from L-aspartic acid or L-glutamic acid were obtained according to the Schotten-Baumann procedure⁸ as shown in Scheme 2. Neutralization of the intermediate diacids, under nitrogen atmosphere and at 0°C, by two equimolar amounts of sodium hydroxyde gave the corresponding di-sodium salts. The latter were recovered by lyophilization in 70% yield⁹ with aspartic acid, compound 4m; and in 90% yield¹⁰ with glutamic acid, compound 5m, respectively.

Scheme 2
$$H_2N$$
 (CH₂)_n-CO₂H $\stackrel{1)}{\longrightarrow}$ (CH₂)₈ $\stackrel{O}{\longrightarrow}$ CO₂ /Na⁺ (CH₂)_n-CO₂ /Na⁺ $\stackrel{O}{\longrightarrow}$ (CH₂)_n $\stackrel{O}{\longrightarrow}$

3-Compounds derived from cysteinic acid and from L-serine: the compound 6m was prepared by reacting undec-10-enoyl chloride with L-serine using a temporary protection of the carboxylic function by a trimethylsilyl ester¹¹. Sulfation of the alcohol function of the amino-acid lateral chain by chlorosulfonic acid in pyridine^{12,13}, followed by stechiometric neutralization with sodium hydroxyde, gave the monomer 6m in 48% yield¹⁴ (Scheme 3).

N-acylation of cysteinic methyl ester, in biphasic medium (aqueous sodium carbonate / chloroform), by undec-10-enoyl chloride, followed by a saponification reaction, gave compound 7m in 90% yield¹⁵ after lyophilization. Due to the fact that both reactions of the acid chloride according to the Schotten-Baumann procedure or with the temporary protection failed, the protection of the methyl ester has proven to be necessary.

Trianionic compound

The structure of this compound is shown in Scheme 4. The intermediate trinitrile 8 was obtained from the tris (hydroxymethyl) aminomethane by a Michaël addition ^{16,17}. This nitrile is successively treated by a methanolic chlorhydric acid solution and by 5N sulfuric acid. Initially, the nitrile function was converted to iminoester which was hydrolyzed by diluted sulfuric acid. The compound 9 was isolated as a colourless oil ^{17,18}.

Triester 9 was reacted with undec-10-enoyl chloride in methylene chloride in the presence of triethylamine to obtain the corresponding amide which, after saponification under sonication, led to the trianionic surfactant 10m isolated with non optimized 40% yield 19.

Tetra-anionic compound

A tetra-anionic surfactant was synthesized by condensing the L-glutamic acid dimethyl ester with the two carboxylic acid functions of the N-(undec-10-enoyl)-L-glutamic acid 11⁴ in the presence of the dicyclohexyl-carbodiimide / N-hydroxysuccinimide couple²⁰ in chloroform (Scheme 5).

The intermediate tetra-ester, isolated after chromatography on silica gel, was saponified by 4N sodium hydroxyde under sonication, neutralized and finally lyophilized to lead to 12m in 80% yield²¹.

Polymerization reaction

Monomeric surfactants 3m to 12m were polymerized by γ -irradiation of a 0.1M aqueous micellar solution according to a previously described procedure³ to lead to polymerized surfactants 3p to 12p. Polyanions thus prepared were purified by gel permeation chromatography (Sephadex-G50) to remove traces of the monomers. The polyanions were recovered by lyophilization. ¹H NMR spectroscopy in D₂O enable us to verify the disappearance of the vinylic protons and the absence of degradation during γ -irradiation.

Biological activity

The different compounds (3p to 12p) were evaluated for their antiviral activity according to well-established procedures³. The compounds were evaluated against HIV in the T4-lymphocyte cell lines CEM-4 and MT-4. (Table 1). In addition, cytotoxicity of the different compounds was determined for the cell lines used. First of all, it is noteworthy that the polyanions were nontoxic for CEM-4 cells as well as for MT-4 cells at concentrations up

400 A. LEYDET et al.

polyanions	CC ₅₀ ^b µg/mL	RT		MTT (CEM-4)		MTT (MT-4)	
		IC ₅₀ ^c μg/mL	SId	IC ₅₀ ^c μg/mL	SI ^d	IC ₅₀ ^c μg/mL	SId
3р	>100	1.7	>58	n.d. ^f	n.d.	4.9	>20
4p	>100	8	>12	n.d.	n.d.	0.8	>125
5p	>100	0.2	>500	0.8	>125	n.d.	n.d.
бр	>100	8	>12	5.9	>17	n.d.	n.d.
7p	>100	8	>12	7.6	>13	n.d.	n.d.
10p	>100	3	>32	3	>32	n.d.	n.d.
12p	>100	30	>3	24	>4	1.1	>90
(CH ₂) ₆ - CO ₂ /N ₆ 13 p ^e	100	3.5	>28	n.d.	n.d.	3.6	>28
Co ₂ : 14pe	770	0.4	1925	0.1	7700	0.2	3850
(CH ₂), NH Y - oso, :15p°	>100	8	>12	11	>9	90	>1
$(CH_{2})_{i} \xrightarrow{O}_{Y : NA^{*}} \begin{cases} co_{i} : 14p^{e} \\ co_{i} : 15p^{e} \\ so_{i} : 16p^{e} \end{cases}$	>100	2	>50	0.8	>125	0.2	>500

Table 1.- Anti-HIV-1 (strain HTLV-III_{B/LAI}) activity of polyanions in CEM-4 and MT-4 cells a

a) All data represent the average value of at least two separate experiments. b) 50% Cytotoxic Concentration, or coumpound concentration required to reduce the viability of uninfected cells by 50% at 5 days of incubation in the presence of the compound. c) 50% Inhibitory Concentration, or compound concentration required to reduce by 50% HIV-1-induced cytopathicity, based on the Reverse Transcriptase (RT) activity or on the MTT assay. d) 50% Selectivity Index or Ratio of CC_{50} to IC_{50} . e) See reference 3. f) not determined.

to 100 μ g/mL. As shown in Table 1, all the polyanions proved inhibitory to HIV-1 in CEM-4 and MT-4 cells. The 50% inhibitory concentration (IC₅₀) based on either the MTT assay³ or reverse transcriptase (RT) activity, fell in the range of 3-24 μ g/mL (MTT) and 0.2-30 μ g/mL (RT), respectively.

The activities of the polyanions 3p, 4p, 6p and 7p, obtained from dianionic surfactants can be compared to those of previously studied polyanions 13p,14p,15p and 16p, derived from analogous monoanionic surfactants in which one carboxylate group is missing (Table 1).

It clearly appears that the introduction of a second anionic group, - i.e. a carboxylate group -, at the end of each hydrocarbon chain, does not significantly change the anti-HIV-1 activity. Likewise, increasing the charge number to three (compound 10p) or four (compound 12p) on each hydrocarbon chain, does not markedly affect the anti-HIV-1 response.

It seems that for this type of polyanions, the anti-HIV-1 activity is not influenced by the local charge density or charge distribution.

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References and Notes

- 1. De Clercq, E.; Clin. Microbiol. Rev., 1995, 8, 200.
- 2. De Clercq, E.; J. Med. Chem., 1995, 38, 2491.
- 3. Leydet, A.; Barthelemy, Ph.; Boyer, B.; Lamaty, G.; Roque, J.P.; Bousseau, A.; Evers, M.; Henin, Y.; Snoeck, R.; Andrei, G.; Ikeda, S.; Reymen, D.; De Clercq, E. J. Med. Chem. 1995, 38, 2433.
- 4. Leydet, A.; El Hachemi, H.; Boyer, B.; Lamaty, G.; Roque, J.P.; Snoeck, R.; Andrei, G.; Ikeda, S.; Reymen, D.; Witvrouw, M.; De Clercq, E. J. Med. Chem. 1995, in press.
- 5. Leydet, A.; Jeantet-Segonds, C.; Barthelemy, Ph.; Boyer, B.; Roque, J.P. Bioorg. Med. Chem. Lett. 1995, submitted.
 - 6. Tomecko, C.G.; Adams, R. J. Am. Chem. Soc. 1927, 49, 522.
 - 7. Compound 3m: Yield = 94%. mp >250°C. IR: KBr, v cm⁻¹: 2928-2856 (C-H); 1590 (C=O, CO_2 -).
- ¹H NMR: (250 MHz, D₂O), δ ppm: 1.3 (m, 14H, H₂⁻ to H₈⁻); 1.7 (q, 2H, H₁⁻, J₂₋₁⁻=7.5Hz, J₂⁻1'=7.5Hz); 2.1 (m, 2H, H9'); 3.1 (t, 1H, H₂, J₂₋₁⁻=7.5Hz); 5.0 (m, 2H, H₁₁⁻, J_{11b-10}⁻ = 18Hz, J_{11a-10}⁻=10.5Hz); 5.8 (m, 1H, H₁₀⁻, J₁₀⁻-11_b⁻=18Hz, J₁₀⁻-11_a⁻=10.5Hz, J₁₀⁻-9⁻=7Hz). MS: (FAB+, G), m/z: 279 (M-Na+2H)+; 301 (M+H)+; 323 (M+Na)+; 623 (2M+Na)+; 923 (3M+Na)+. Anal: C₁₄H₂₂O₄Na₂.
 - 8. a) Schotten, C. Ber. 1884, 17, 2544. b) Baumann, E. Ber. 1886, 19, 3218.
- 9. Compound 4m: Yield = 70%. mp (acide) = 83-85°C. R_f : 0.6 (20% aqueous NH₃/H₂O/EtOH, 16/12/100, v/v/v). [α]_D²⁵ (salt) = +0.8° (H₂O). IR: KBr, v cm⁻¹: 3397 (N-H); 2928-2855 (C-H); 1580-1406 (C=O, CO₂⁻). 1H NMR: (250 MHz, CDCl₃), δ ppm (acide): 1.1 (m, 10H, H₄· to H₈·); 1.4 (m, 2H, H₃·); 1.8 (q, 2H, H₉·, J₉·. 8'=7Hz, J₉·.₁₀'=6Hz); 2.0 (t, 2H, H₂·, J₂·.₃·=7Hz); 2.3 (m, 2H, H₃); 4.4 (q, 1H, H₂, J₂.₃=7Hz, J₂._{NH}=7.5Hz); 4.8 (m, 2H, H₁₁·, J_{11b}·.₁₀·=17.1Hz, J_{11a}·.₁₀·=10Hz); 5.8 (m, 1H, H₁₀·, J₁₀·._{11b}·=17.1Hz, J₁₀·._{11a}·=10Hz); 6.8 (m, 1H, NH); 10.25 (m, 2H, (CO₂H)₂). CNMR: (50.32 MHz, CDCl₃), δ ppm (acid): 25,56 to 36,19 (C₂· to C₉· and C₃); 48.59 (C₂); 114.34 (C₁₁·); 139.08 (C₁₀·); 174.47 (C₁·); 175.04 and 175.27 (C₁ and C₄). MS: (FAB+, NBA), acid, m/z: 322 (M+Na)+; 300 (M+H)+. Anal: C₁₅H₂₃NO₅Na₂.
- 10. Compound 5m: Yield = 90%. mp (acid) = 77-79°C. R_f : 0.5 (20% aqueous NH₃/H₂O/EtOH, 16/12/100, v/v/v).[α]_D²⁵ (salt) = +0.2° (H₂O). IR: KBr, v cm⁻¹: 3392 (N-H); 2928-2853 (C-H); 1580-1409 (C=O, CO₂⁻).

 1H NMR: (250 MHz, CDCl₃), δ ppm, (acid): 1.2 (m, 12H, H₄· to H₈· and H₃); 1.4 (m, 2H, H₃·); 1.7 (q, 2H, H₉·, J₉·-₈·=7Hz, J₉·-₁₀·=6Hz); 2.1 (t, 2H, H₂·, J₂·-₃·=7Hz); 2.3 (m, 2H, H₄); 4.4 (q, 1H, H₂, J₂-₃-7Hz, J₂-NH=7,5Hz); 4.8 (m, 2H, H₁₁·, J_{11b}·-₁₀·=17.1Hz, J_{11a}·-₁₀·=10Hz); 5.8 (m, 1H, H₁₀·, J₁₀·-_{11b}·=17.1Hz, J₁₀·-_{11a}·=10Hz); 6.8 (m, 1H, NH); 10.3 (m, 2H, (CO₂H)₂).¹³C NMR: (50.32, CDCl₃), δ ppm (acid): 25,64 to 36,29 (C₂· to C₉· and C₃, C₄); 51.77 (C₂); 114.21 (C₁₁·); 139.11 (C₁₀·); 175.04 and 175.19 (C₁ and C₅): 177.41 (C₁·). MS: (FAB⁺, NBA), (acid), m/z: 322 (M+Na)⁺; 300 (M+H)⁺. Anal: C₁₆H₂₅NO₅Na₂.
 - 11. Schwabacher, A.W.; Lei, H. J. Org. Chem. 1990, 55, 6080. See also reference 4.
 - 12. Neuberg, C.; Liebermann, L. Biochem. Z. 1921, 121, 326.

A. LEYDET et al.

- 13. Tipson, R.S. "Sulfonic Esters of Carbohydrates", Advances in Carbohydrate Chemistry, 1953, 8, 107.
- 14. Compound 6m: Yield = 48%. mp > 250°C. Rf: 0.43 (AcOEt/MeOH, 5/5,v/v). $[\alpha]_D^{25} = +15.3^\circ$ (c = 0.011, H₂O). IR: KBr, v cm⁻¹: 3335 (N-H); 2921-2855 (C-H, CH₂); 1637 (C=O, amide); 1616 (C=O, CO₂⁻); 1140 (S=O, sulfate). ¹H NMR: (250 MHz, D₂O), δ ppm: 1.37 (m, 10H, H₄· to H₈·); 1.54 (m, 2H, H₉·); 2.08 (m, 2H, H₃·); 2.23 (t, 2H, H₂·, J₂·-3·=7.6Hz); 3.22 (m, 2H, H₃); 4.45 (m, 1H, H₂); 4.93 (m, 2H, H₁₁·, J_{11b}·. 10° =17.4Hz, J_{11a}·-10·=10.4Hz); 5.87 (m, 1H, H₁₀·, J₁₀·-11b·=17.4Hz, J₁₀·-11a·=10.4Hz).
- 13C NMR : (50.32, D₂O), δ ppm: 26.0 (C_{3'}); 28.94 to 29.19 (C_{4'} to C_{8'}); 33.90 (C_{9'}); 36.63 (C_{2'}); 55.12 (C₂); 69.55 (C₃); 114.77 (C_{11'}); 141.34 (C_{10'}); 175.76 (C₁'); 177.59 (C₁). MS : (FAB⁻, TG), m/z : 350 (A²⁻+H)⁻; 372 (A²⁻+Na)⁻; 767 (2A-2H+3Na)⁻. Anal : C₁₄H₂₃NO₇SNa₂.
- 15. Compound 7m: Yield = 90 %. mp > 250°C. R_f : 0.43 (AcOEt/MeOH, 5/5, v/v). $[\alpha]_D^{25} = -9.07^\circ$ (c = 0.105, H₂O).IR : KBr, v cm⁻¹: 3346 (N-H); 2921-2844 (C-H, CH₂); 1643 (C=O, amide); 1599.7 (C=O, CO₂⁻); 1044 (S=O, sulfonate). ¹H NMR : (250 MHz, D₂O), δ ppm: 1.25 (m, 10H, H₄· to H₈·); 1.67 (t, 2H,H₉·); 2.11 (m, 2H, H₃·); 2.38 (t, 2H, H₂·, J_{2·-3}·=7Hz); 4.35 (m, 2H, H₃); 4.55 (m,1H, H₂); 5,05 (m, 2H, H₁₁·, J_{11b·.} 10·=17.4Hz, J_{11a·-10}·=10.4Hz); 6.0 (m, 1H, H₁₀·, J_{10·-11b}·=17.4Hz, J_{10·-11a}·=10.4Hz). ¹³C NMR : (50,32, D₂O), δ ppm: 25.85 (C₃·); 28.92 to 29.18 (C₄· to C₈·); 33.89 (C₉·); 36.69 (C₂·); 52.66 (C₃); 52.82 (C₂); 114.76 (C₁₁·); 141.37 (C₁₀·); 177.2 (C₁·); 177.31 (C₁). MS : (FAB-, TG), m/z : 350 (A²⁻+H)-; 372 (A²⁻+Na)-; 767 (2A-2H+3Na)-. Anal : C₁₄H₂₃NO₆SNa₂.
 - 16. Newkome, G.R.; Moorefield, C.N.; Theriot, K.J. J. Org. Chem. 1988, 53, 5552.
 - 17. Newkome, G.R.; Lin, X. Macromolecules 1991, 24, 1443.
 - 18. Dayan, I.; Libman, J.; Shanzer, A.; Felder, L.E. J. Am. Chem. Soc. 1991, 113, 3431.
- 19. Compound 10m: Yield = 40%, mp =182-184°C.IR: KBr, v cm⁻¹: 3474-3415 (N-H); 2927-2854 (C-H); 1639 (C=O amide); 1573 (C=O, CO₂-). ¹H NMR: (250 MHz, D₂O), δ ppm: 1.2 (m, 10H, H₄· to H₈); 1.5 (m, 2H, H₃·); 2.0 (q, 2H, H₉·, J₉₋₈=6.8Hz, J₉₋₁₀=6.8Hz); 2.1 (t, 2H, H₂·, J₂₋₃=8Hz); 2.4 (t, 6H, H₂, J₂₋₃=6.8Hz); 3.7 (t, 12H, H₃, H₄, J₂₋₃=6.8Hz); 4.9 (m, 2H, H₁₁·); 5.8 (m, 1H, H₁₀·). MS: (FAB+, NOBA), m/z: 592 (M+Na)⁺; 570 (M+H)⁺; 548 (M-Na+2H)⁺; 526 (M-2Na+3H)⁺. Anal: C₂₄H₃₈NO₁₀Na₃.
 - 20. Anderson, G.W.; Zimmerman, J.E.; Callahan, F.M. J. Am. Chem. Soc. 1964, 86, 1839.
- 21. Compound 12m: Yield = 80%. mp >260°C.[α]D²⁵ = -0.16° (H₂O). IR: KBr, v cm⁻¹: 3396 (N-H); 2928-2855 (C-H); 1580 (C=O, CO₂⁻). ¹H NMR: (250 MHz, D₂O), δ ppm: 1.2 (m, 12H, H₄· to H₈· and H₃); 1.5 (m, 2H, H₃·); 2.0 (m, 6H, H₉·, H₁₃, H₈); 2.2 (m, 6H, H₂·, H₃·, H₉); 2.4 (m, 4H, H₄, H₁₄); 4.3 (t, 1H, H₇, J₇. 8=7Hz); 4.4 (m, 2H, H₁₂); 4.8 (m, 2H, H₁₁·, J_{11b'-10'}=17Hz, J_{11a'-10'}=10Hz); 6.0 (m, 1H, H_{10'}, J_{10'-11b'}=17Hz, J_{10'-11a'}=10Hz, J_{10'-11a'}=6.5Hz). ¹³C NMR: (50.32, D₂O), δ ppm: 26.2 (C_{3'}); 29.1 to 32.8 (C₃· to C_{8'} and C₃, C₁₃); 34.0 (C_{9'}); 35.2 (C₄, C₁₄); 36.6 (C_{2'}); 54.0 (C₇); 56.3 (C₂, C₁₂); 115.0 (C_{11'}); 141.6 (C_{10'}); 173.7 to 178.2 (C_{1'}, C₆ and C₁₀); 179.5 to 183.0 (C₁, C₅, C₁₁ and C₁₅). MS: (FAB⁻, Glycerol), m/z: 636 (M-Na)⁻; 658 (M-H)⁻; 614 (M-2Na+H)⁻. Anal: C₂₆H₃₇N₃O₁₁Na₄.

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